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**PROFESSIONAL BACKGROUND**

***Academic Positions***

**January 2019 - current:** Director of CSELS, Center for the Science and Engineering of Living Systems, Washington University in St. Louis

**March 2015 – current:** Edwin H. Murty Professor of Engineering, McKelvey School of Engineering, Washington University in St. Louis

**September 2012 – December 2018:** Director, Center for Biological Systems Engineering, Washington University in St. Louis, School of Engineering & Applied Sciences

**January 2013 – January 2018:** Co-Director, Center for High Performance Computing, Washington University in St. Louis

**January 2011 – current:** Professor, Department of Biomedical Engineering, Washington University in St. Louis, School of Engineering & Applied Sciences

**March 2007 – current:** Member, Hope Center for Neurodegenerative Disorders, Washington University School of Medicine

**July 2007 – December 2010:** Associate Professor, Department of Biomedical Engineering, Washington University in St. Louis, School of Engineering & Applied Sciences; Adjunct Associate Professor, Department of Biochemistry & Molecular Biophysics, Washington University School of Medicine

**February 2009 – August 2010:** Director, Center for Computational Biology, Washington University School of Medicine

**May 2002 – August 2010:** Member, Center for Computational Biology, Washington University School of Medicine

**September 2001 – June 2007:** Assistant Professor, Department of Biomedical Engineering, Washington University in St. Louis, School of Engineering & Applied Sciences; Adjunct Assistant Professor, Department of Biochemistry and Molecular Biophysics, Washington University School of Medicine

***Non-academic Affiliations***

**January 2019 - current:** Member, Scientific Advisory Board, Dewpoint Therapeutics, Boston, MA and Dresden, Germany

## ***Education and Training***

### **Postdoctoral Scientist, 1998-2001**

Department of Biophysics and Biophysical Chemistry, Johns Hopkins University, School of Medicine, Baltimore, MD; Mentor: Professor George D. Rose

### **Postdoctoral Scientist, 1996-1998**

Department of Biochemistry and Molecular Biophysics, Washington University in Saint Louis, School of Medicine, St. Louis, MO; Mentor: Professor Jay W. Ponder

### **Ph.D., Biological Physics, 1992-1996**

Department of Physics and Astronomy, Tufts University, Medford, MA

Advisor: Professor David L. Weaver (deceased)

Thesis: Algorithms for modeling folding pathways of proteins.

### **M.S., Solid State Physics, 1990-1992**

Department of Physics and Astronomy, Tufts University, Medford, MA

### **B.Sc., Honors in Physics, Mathematics, and Electronics, 1986-1989**

St. Joseph's College, Bangalore University, Bangalore, India

## **RESEARCH PROGRAM**

### ***Summary of research interests***

Research in my lab is focused on uncovering the biophysical principles underlying the form, function, regulation, phase behavior, and evolution of intrinsically disordered and multivalent proteins. These foci directly impact topics of biomedical relevance that include the molecular basis of neurodegeneration, phase transitions that lead to protein and RNA condensates, the biophysics of molecular recognition via soft and disordered interfaces, and the design of responsive, protein-based biomaterials. Our work is driven by a blend of novel multiscale computer simulations, adaptations and developments of polymer physics theories, the physics of active matter and non-equilibrium statistical physics of living systems. We close the loop with *in vitro* and in cell experiments performed in my lab and through a vibrant network of international collaborations.

**Intrinsically disordered proteins (IDPs):** Between 30% and 50% of eukaryotic proteomes comprise of intrinsically disordered proteins / regions (IDPs / IDRs). Over the years, we have uncovered sequence-ensemble relationships of IDPs / IDRs that connect the information encoded in amino acid sequences to quantitative descriptions of conformational ensembles. These efforts have yielded predictive descriptions and uncovered the rich contributions made by amino acid composition, charge content, and sequence patterning effects to the global sizes, shapes, amplitudes of conformational fluctuations, and context dependent secondary structural preferences within IDPs / IDRs. These discoveries have been made possible through novel combinations of polymer physics theories, accurate and efficient molecular simulations driven by our homegrown ABSINTH implicit solvation model, and biophysical experiments. In ongoing investigations, we are pursuing three new avenues: (1) We are using computationally driven approaches to design novel IDPs / IDRs with bespoke sequence-ensemble relationships to uncover the connections between these relationships and molecular functions, especially in the context of transcriptional regulation. (2) We are developing and deploying new, multiscale computational and theoretical approaches to model circuits controlled by multisite

phosphorylation and other multisite post-translational modifications within IDPs / IDRs. (3) We are deploying new methods to investigate the effects of sequence-specific charge regulation, specifically proton uptake and release phenomena on disorder-order transitions of IDPs / IDRs in cellular milieus.

**Phase transitions in cell biology:** We are actively working on the problem of phase transitions that are controlled or influenced by multivalent proteins and RNA molecules. These phase transitions include sol-gel transitions as well as liquid-liquid, liquid-solid, and liquid-liquid crystalline phase separation. Our focus is on describing relationships between protein sequences / architectures and the stimulus specific phase diagrams that are governed by spontaneous driving forces also known as passive processes. This focus is driven by the importance of biomolecular condensates that form via phase transitions afford spatiotemporal organization and information transduction within cells. Specific foci include nuclear bodies, stress responses, synaptic bodies, and the interplay between spontaneous and driven processes. We are also embarking on rigorous dynamical and structural descriptions of biomolecular condensates with a view to uncovering how driven (active) processes influence spatiotemporal regulation in living cells.

**Neurodegeneration:** We work on connecting the driving forces for and the mechanisms of polyglutamine aggregation and phase separation to intracellular interactions that lead to neurodegeneration in HD and other polyglutamine expansion disorders. An emerging focus is on the modulation of aggregation and phase behavior by endogeneous networks of protein-protein interactions.

**Intrinsically disordered regulome in bacteria:** Although IDPs / IDRs represent only 5% of typical bacterial proteomes, it has become clear that almost all key aspects that control bacterial life cycles are controlled by IDPs / IDRs. Often these sequences are hypervariable and the implications an emerging goal in my lab is to uncover how IDPs / IDRs influence functional outcomes in bacteria. These studies are being driven by multilevel theory, computation, biochemical experiments, and high-throughput library-based in vivo investigations. The long-term goal is to combine bottom-up and top-down investigations to make headway into the design of bacterial communities with bespoke emergent properties and the selective inhibition of bacteria by targeting IDPs / IDRs using novel antibiotics.

**Molecular and Cellular Engineering:** We are building on our work pertaining to phase transitions and IDPs to develop, prototype, and deploy computational methods to predict phase behavior from amino acid sequence and advance the design of responsive peptide and protein-based biomaterials.

### ***Professional Societies***

Member, American Association for the Advancement of Science (AAAS)

Member, American Chemical Society (ACS)

Member, American Institute of Medical and Biological Engineers (AIMBE)

Member, Biophysical Society (BPS) and subgroups on Intrinsically Disordered Proteins (IDPs) and Biopolymers In Vivo (BIV)

Member, The Protein Society (TPS)

Member, by nomination, Sigma Xi, The Scientific Honor Society

### ***Awards***

Recipient of the 2019 McKelvey Engineering Faculty Award for excellence in teaching, Washington University in St. Louis, April 2019

Named as a Mercator Fellow by the Deutsche Forschungsgemeinschaft to serve as an international collaborator with Professor Dr. Edward A. Lemke, Professor Dr. Simon Alberti, and Professor Dr. Claus A.M. Seidel.

Elected Fellow of the Biophysical Society, 2019

Dean's Award for Outstanding Contributions in Service of the Mission of the School of Engineering & Applied Sciences, Washington University in St. Louis, April 2017.

Inducted to the College of Fellows of the American Institute for Medical and Biological Engineering (AIMBE), April 2016.

Elected fellow of the American Association for Advancement of Science (AAAS), November 2013

Basil O'Connor Starter Scholar Award, March of Dimes Research Foundation, 2004

Graduate Biophysics Fellowship, selected to attend Princeton-NEC workshop, June 1996

John F. Burlingame Graduate Research Fellowship, Tufts University, 1995 – 1996

National Merit Scholarship, Bangalore University, Bangalore, India, 1989

### **Honors**

Listed, in 2019, by the Institute of Science as being in the top 1% of researchers worldwide in terms of number of citations

Chair of NIH ZRG1-MDCN study section, April 2019

Invited participant and speaker in the Banbury Center meeting on Phase Separated Assemblies in Cell Biology, Cold Spring Harbor Laboratory, December 16-19, 2018

Co-Chair of NIH ZRG1-MDCN study section, July 2018

Member, Scientific Advisory Board at the Max Planck Institute of Molecular Cell Biology and Genetics, Dresden, Germany, term ending December 31, 2022

Chair the 2017 FASEB Summer Research Conference on Molecular Mechanisms and Physiological Consequences of Protein Aggregation

Guest Editor, *Current Opinion in Structural Biology*, 2017

Guest Editor, Special Issue of *Seminars on Cell and Developmental Biology*, 2015

Member, Editorial Advisory Board, *Biophysical Journal*, December 2014 – current

Member, Executive Committee, Protein Folding Consortium, Sponsored by the National Science Foundation, Research Coordination Network, September 2014 – current

Member, Editorial Board of *Protein Engineering, Design and Selection*, July 2014 – current

Elected Symposium Co-Chair for the 9<sup>th</sup> Annual Symposium of the Biophysical Society's Intrinsically Disordered Proteins Subgroup, February 2014

Faculty of 1000 Member, *Structural Biology: Theory & Simulation Section*, April 2013 – August 2016

BMC Biophysics, Editorial Board Member, 2012 – current.

Chair, 2<sup>nd</sup> Gordon Research Conference on Intrinsically Disordered Proteins, 2012

One of Fifteen Scientists Invited by the National Science Foundation to Participate in a Workshop to Discuss the Future and Frontiers of Protein and RNA Biophysics, September 2011

Charter Member, National Institutes of Health Center for Scientific Review, Biophysics of Neural Systems Study Section, June 2008 – June 2012

Member, Protein Folding Consortium, supported by the National Science Foundation, Research Coordination Network, May 2009 – current

Chair, by election, Intrinsically Disordered Proteins Subgroup, Biophysical Society, 2009

Regular Member, National Science Foundation, Molecular & Cellular Biosciences, Biophysics Panel, April 2008 – Current

### ***Named / Keynote Lectures and Webinars***

Closing Keynote Speaker, EMBO workshop on *Intrinsically Disordered Proteins: From Molecules to Systems*, Bangalore, India, December 2019.

Featured speaker, American Chemical Society Webinar Series, Phase Separation of Multivalent Proteins, co-produced with Biochemistry, November 2019

Opening Keynote Speaker, Gordon Research Conference on Computational Aspects of Biomolecular NMR, Les Diablerets, Switzerland, June 2019.

Student Invited Closing Keynote Speaker for Gordon Research Seminar on Computational Aspects of Biomolecular NMR, Les Diablerets, Switzerland, June 2019: *Uncovering the marvels of protein conformational heterogeneity along roads less traveled.*

Keynote Speaker, Dutch Biophysical Society, Veldhoven, Netherlands, October 2018.

Invited to give the Telluride town talk entitled *Neurodegeneration from the ground up* as an accompaniment to the Telluride Science Research Conference Series, July 2015.

Keynote Distinguished Speaker, Syracuse Biomaterials Institute, Syracuse University, April 2017.

Keynote Speaker, International Symposium on Chromatin Dynamics: Theoretical and Polymer Physics Approaches, Hiroshima, Japan, December 2015.

Opening Plenary Speaker, 12<sup>th</sup> Annual New England Structure Symposium (NESS), Theme: Structure and Dynamics of Intrinsically Unfolded Proteins, sponsored by the University of Connecticut, Storrs, CT, October 2015.

Keynote Speaker, 29<sup>th</sup> Annual Gibbs Conference on Biothermodynamics, Touch of Nature Conference Center, Carbondale, IL October 2015.

Keynote Speaker at the Annual Symposium of the Cellular Dynamics and Macromolecular Complexes CREATE Graduate Training Program, University of Montreal, Canada, August 2015.

Keynote Speaker, Intrinsically Disordered Proteins Gordon Research Conference, July 2014.

Keynote Speaker, 8<sup>th</sup> Annual Symposium of the Biophysical Society's Intrinsically Disordered Proteins Subgroup, February 2014.

Keynote Speaker, Annual Midwest Conference on Protein Folding, Assembly, and Molecular Motions, May 2011.

### ***Invited Talks at National and International Conferences / Symposia / Workshops***

Invited Speaker, Cell Symposia on *Biological Assemblies: Phase separation and more*, Seville, Spain, October 2020.

Invited Speaker, NSF Sponsored Workshop and International Summer School on *Genome Architecture and Dynamics*, Massachusetts Institute of Technology, Cambridge, MA, June 2020.

Invited Speaker, Royal Society of Chemistry sponsored workshop on *Anhydrous Proteins*, Chichley Hall, Buckinghamshire, UK, March 2020.

- Invited Speaker, *Telluride Science Research Conference on Phase Separation in Biology*, Telluride, CO, March 2020.
- Invited Speaker, *International Symposium on Membraneless Organelles in Cell Life and Disease*, Seville, Spain, March 2020.
- Invited Speaker, 2019 Symposium on Intrinsically Disordered Proteins, Stanford University, February 2020.
- Invited Speaker, Telluride Science Research Conference on *Intrinsically Disordered Proteins*, July 2019.
- Invited Speaker, *Cell Press Symposium on Regulatory RNAs*, Berlin, Germany, May 2019.
- Invited Speaker, Keystone Symposium on *Biomolecular Condensates: Phase separated organizers of cellular biochemistry*, Snowbird, Utah, April 2019.
- Invited Symposium Speaker, 63<sup>rd</sup> Annual Meeting of the Biophysical Society, Baltimore, March 2019.
- Invited Speaker, *Symposium on Phase Separation in Biology and Disease*, New York Academy of Sciences, February 2019.
- Invited Speaker, *workshop on Phase Transitions in Polymeric and Protein Systems*, Organized by the Max Planck Institute for Physics of Complex Systems, Dresden, Germany, February 2019.
- Invited Lecturer, *EMBO course on Phase Transitions in Polymeric and Protein Systems*, Organized by the Max Planck Institute for Cell Biology & Genetics, Dresden, Germany, February 2019.
- Invited Speaker, 2019 *BMES Cellular and Molecular Bioengineering Conference*, San Diego, CA, January 2019.
- Invited Speaker and Session Chair, *Intrinsically Disordered Proteins Gordon Research Conference*, Les Diablerets, Switzerland, July 2018.
- Invited Speaker, *EMBO | EMBL Symposium: Cellular Mechanisms Driven by Liquid Phase Separation*, Heidelberg, Germany, May 2018.
- Invited Speaker, *ASBMB Symposium at Experimental Biology*, San Diego, CA, April 2018.
- Invited Speaker, 13<sup>th</sup> *CHDI Annual Huntington's Disease Therapeutics Conference: A forum for Drug Discovery and Development*, Palm Springs, CA, February 2018.
- Invited Speaker, *International Conference on Intrinsically Disordered Proteins*, IISER Mohali, India, December 2017.
- Invited Speaker, Build the Cell Subgroup Symposium at the Annual Meeting of the American Society for Cell Biology, Philadelphia, PA, December 2017.
- Invited Speaker, Telluride Science Research Conference on Intrinsically Disordered Proteins and Membraneless Organelles, Telluride, CO, July 2017.
- Invited Speaker, Proteins Gordon Research Conference, Holderness, New Hampshire, June 2017.
- Invited Speaker, Inaugural symposium of the St. Jude Research Collaborative on membraneless organelles, The Biology of Liquid Organelles, St. Jude Children's Research Hospital, May 2017.
- Invited Speaker, New Frontier in Cellular Structure and Function, Genetics, Genomics and Systems Biology Symposium Sponsored by the University of Chicago, May 2017.

- Invited Speaker, VIB Conference Series, Phase Transitions in Biology and Disease, Leuven, Belgium, May 2017.
- Invited Speaker, Symposium on Coacervation: Physics, Chemistry, and Biology. Part of the 253<sup>rd</sup> National Meeting of the American Chemical Society, San Francisco, April 2017.
- Invited Speaker, 3<sup>rd</sup> International Workshop on Protein Folding & Dynamics, National Centre for Biological Sciences, Bangalore, India, November 2016.
- Invited Speaker, 2<sup>nd</sup> COST Symposium on Non-Globular Proteins in Molecular Pathophysiology, Belgrade, Serbia, September 2016.
- Invited Speaker, Physical Chemistry Division Symposium In Intrinsically Disordered Proteins at the 252<sup>nd</sup> National Meeting of the American Chemical Society, August 2016.
- Invited Speaker, 30<sup>th</sup> Anniversary Symposium of the Protein Society, July 2016.
- Invited Speaker, 3<sup>rd</sup> Workshop on the Physical Basis of Cellular Adaptation & Memory, Bellairs Research Institute, Barbados, April 2016.
- Invited Speaker, CHDI Htt Protein Lifecycle Workshop, New York City, March 2016.
- Invited Symposium Speaker, 60<sup>th</sup> Annual Meeting of the Biophysical Society, Los Angeles, CA March 2016.
- Invited Speaker, Southeast and Southwest regional meeting of the American Chemical Society, Symposium on Intrinsically Disordered Proteins, Memphis, TN, November 2015.
- Invited Speaker, CECAM Workshop on Computational Modeling of Intrinsically Disordered Proteins, Zurich, Switzerland August 2015.
- Invited Speaker, FASEB meeting on Molecular Mechanisms and Physiological Consequences of Protein Aggregation, June 2015.
- Invited Speaker, Proteins Gordon Research Conference, Holderness, New Hampshire, June 2015.
- Invited Speaker, Conference on Intracellular Phase Transitions: RNA, Protein, Lipids, and Beyond. Princeton University, April 2015.
- Invited Speaker, International Scientific Seminar, Chromosome Dynamics: Computational Models and Experimental Data, Sponsored by the Royal Society, Chicheley Hall, Buckinghamshire, UK, November 2014.
- Invited Speaker and Session Chair, Thematic Meeting, Disordered Motifs and Domains in Cell Control, Dublin Ireland – Sponsored by the Biophysical Society, October 2014.
- Invited Speaker, Gordon Research Conference on Protein Folding Dynamics, January 2014.
- Invited Speaker, 3<sup>rd</sup> USA-Mexico Workshop in Biological Chemistry: Protein Folding, Dynamics, and Function, November 2013.
- Invited Speaker, CECAM International Workshop on Intrinsically Disordered Proteins, ETH Zurich, September 2013.
- Invited Speaker, FASEB Summer Research Conference on Molecular Mechanisms and Physiological Consequences of Protein Aggregation, June 2013.
- Invited Speaker, 57<sup>th</sup> Annual Meeting of the Biophysical Society, February 2013.
- Invited Speaker, International Symposium on Protein Folding, National Centre for Biological Sciences, Bangalore, India, October 2012

Invited Speaker, Biopolymers Gordon Research Conference, June 2012

Invited Speaker, Gordon Research Conference on Protein Folding Dynamics, January 2010

Invited Speaker, FASEB Summer Research Conference on Biophysics and Biology of Amyloids, June 2009

Invited Speaker and Session Chair, 53<sup>rd</sup> Annual Meeting of the Biophysical Society, March 2009

Invited Speaker, 22<sup>nd</sup> Annual Symposium of The Protein Society, July 2008

Invited Speaker, Biopolymers Gordon Research Conference, June 2008

Invited Speaker and Symposium Co-Chair, 2<sup>nd</sup> Annual Symposium of the Biophysical Society's Intrinsically Disordered Proteins Subgroup, February 2008

Invited Speaker, FASEB Amyloid Meeting, June 2006

Invited speaker, I2CAM Workshop on Protein Aggregation and Amyloid Formation in Systemic and Neurodegenerative Diseases, EPFL, Lausanne, Switzerland, July 2005

**Peer Reviewed Publications (reverse chronological order) includes articles in press (Google scholar h-index: 50; i10 index 96)**

1. J. A. Greig, T.A. Nguyen, M. Lee, A. S. Holehouse, A.E. Posey, **R.V. Pappu**, G.J. Jedd. Arginine-enriched mixed-charge domains provide cohesion for nuclear speckle condensates. Accepted for publication in *Molecular Cell*.
2. J-M. Choi, A.S. Holehouse, **R.V. Pappu**. Physical principles underlying the complex biology of intracellular phase transitions. *Annual Review of Biophysics, in press*.
3. J-M. Choi<sup>†</sup>, F. Dar<sup>†</sup>, **R.V. Pappu**. LASSI: A lattice model for simulating phase transitions of multivalent proteins. (<sup>†</sup>Co-first authors). *PLoS Computational Biology*, **15**: e1007028.
4. M. J. Fossat, **R.V. Pappu**. (2019). *q*-Canonical Monte Carlo sampling for modeling the linkage between charge regulation and conformational equilibria of peptides. *Journal of Physical Chemistry B*, **123**: 6952-6967.
5. M.C. Cohan, K.M. Ruff, **R.V. Pappu**. (2019). Information theoretic measures for quantifying sequence-ensemble relationships of intrinsically disordered proteins. *Protein Engineering, Design, and Selection, in press* <https://doi.org/10.1093/protein/gzz014>.
6. S.K. Powers, A.S. Holehouse, D.A. Korasick, K.H. Schreiber, E. Tycksen, J.M. Jez, **R.V. Pappu**, L.C. Strader. (2019). Nucleo-cytoplasmic partitioning of ARF proteins control auxin responses in *Arabidopsis thaliana*. *Molecular Cell*, **76**: 175-190.e5.
7. W. Bai, C. Sargent, J-M. Choi, **R.V. Pappu**, F. Zhang. (2019). Covalently assembled single chain protein nanostructures with ultrahigh stability. *Nature Communications*, **10**: 3317.
8. J. Cable, C. Brangwynne, G. Seydoux, D. Cowburn, **R.V. Pappu**, C.A. Castañeda, L.E. Berchowitz, Z. Chen, M. Jonikas, A. Dernburg, T. Mittag, N.L. Fawzi. Phase separation in Biology and Disease – a symposium report. (2019). *Annals of the New York Academy of Sciences, in press*. <http://dx.doi.org/10.1111/nyas.14126>.
9. I. Peran\*, A.S. Holehouse\*, I.S. Caricco, **R.V. Pappu**<sup>†</sup>, O. Bilsel<sup>†</sup>, D.P. Raleigh. (2019). Unfolded states under folding conditions accommodate sequence-specific conformational preferences alongside ensemble-averaged features of random coils. *Proceedings of the National Academy of Sciences USA*. **116** (25): 12301 – 12310. (\*Co-first authors; <sup>†</sup>Co-corresponding authors).

10. S. Boeynaems<sup>†</sup>, A.S. Holehouse, V. Weinhardt, D. Kovacs, J. Van Lindt, C. Larabell, L. Van Den Bosch, R. Das, P. Tompa, **R.V. Pappu**<sup>†</sup>, A. Gitler<sup>†</sup>. (2019). Spontaneous driving forces give rise to protein-RNA condensates with coexisting phases and complex material properties. *Proceedings of the National Academy of Sciences USA*. **116** (16): 7889-7898. (<sup>†</sup>Co-corresponding authors).
11. R. Beveridge\*, L. G. Migas\*, R. K. Das, **R. V. Pappu**, R. W. Kriwacki, P. E. Barran. (2019). Ion mobility mass spectrometry helps uncover the impact of charge patterning on the conformational distributions of intrinsically disordered proteins. *Journal of the American Chemical Society*, **141**(12): 4908-4918. (\*Co-first authors).
12. J-M. Choi, **R.V. Pappu**. (2019). Improvements to the ABSINTH forcefield for proteins based on experimentally derived amino-acid specific backbone conformational statistics. *Journal of Chemical Theory and Computation*, **15**: 1367-1382.
13. J-M. Choi, **R.V. Pappu**. (2019). Experimentally derived and computationally optimized conformational statistics for blocked amino acids. *Journal of Chemical Theory and Computation*, **15**: 1355-1366.
14. K. M. Ruff\*, **R.V. Pappu**\*, A.S. Holehouse\*. (2019). Conformational preferences and phase behavior of low complexity sequences: Insights from multiscale simulations. *Current Opinion in Structural Biology*, **56**: 1-10. (\*Co-corresponding authors).
15. A.E. Posey<sup>†</sup>, A.S. Holehouse<sup>†</sup>, **R.V. Pappu**. (2018). Phase separation of intrinsically disordered proteins. *Methods in Enzymology*, **611**: 1-30. (<sup>†</sup>Co-first authors).
16. S. Roberts, T. S. Harmon, J. Schaal, K. Li, A. Hunt, V. Miao, Y. Wen, T. G. Oas, J. Collier, **R. V. Pappu**, A. Chilkoti. (2018). Modulation of order and disorder in recombinant polypeptides creates injectable tissue integrating networks. *Nature Materials*, **17**: 1154–1163. **In the news:** <https://pratt.duke.edu/about/news/biomaterials-frankenstein-proteins-help-heal-tissue>
17. G. Fuertes, N. Banterle, K.M. Ruff, A. Chowdhury, **R.V. Pappu**\*, D. I. Svergun\*, E.A. Lemke\*. (2018). Comment on “Innovative scattering analysis shows that hydrophobic disordered proteins are expanded in water.” *Science*, **361**: eaau8230. Technical comment solicited by the editors. (\*Co-corresponding authors).
18. T.Y. Yoo, J-M. Choi, W. Conway, C-H. Yu, **R.V. Pappu**, D.J. Needleman. (2018). Measuring NDC80 binding reveals molecular basis of tension-dependent kinetochore-microtubule attachments. *eLife*, **7**: e36392.
19. K.M. Ruff, S. Roberts, A. Chilkoti\*, **R.V. Pappu**\*. (2018). Advances in understanding stimulus responsive phase behavior of intrinsically disordered protein polymers. *Journal of Molecular Biology*. **430**: 4619-4635. (\*Co-corresponding authors)
20. A.E. Posey, **R.V. Pappu**. (2018). A first glimpse of nucleation of phase transitions in living cells. (*Invited preview*). *Molecular Cell*, **71**: 1-3.
21. J. Wang, J-M. Choi, A.S. Holehouse, X. Zhang, M. Jahnel, S. Maharana, R. Lemaitre, A. Pozniakovski, D. Drechsel, I. Poser, **R. V. Pappu**, S. Alberti, A. A. Hyman. (2018). A molecular grammar underlying the driving forces for phase separation of prion-like RNA binding proteins. *Cell*, **174**: 688-699.
22. H. Mirbaha, D. Chen, O.A. Morozova, K.M. Ruff, A. Sharma, X. Liu, M. Goodarzi, **R.V. Pappu**, D.W. Colby, H. Mirzai, L.A. Jochimiak, M.I. Diamond. (2018). Inert and seed-competent tau monomers elucidate the structural origins of aggregation. *eLife*, **7**: e36584.
23. A. Mittal, A.S. Holehouse, M.C. Cohan, **R.V. Pappu**. (2018). Sequence-to-conformation

- relationships of disordered regions tethered to folded domains of proteins. *Journal of Molecular Biology*, **430**: 2403-2421.
24. E. A. Newcombe<sup>§</sup>, K. M. Ruff<sup>§</sup>, Y. M. Ramdhan, A. Sethi, A. R. Ormsby, O. Kleifeld, A. W. Purcell, P. R. Gooley, **R. V. Pappu**<sup>\*</sup>, D. M. Hatters<sup>\*</sup>. (2018). Tadpole-like conformations of huntingtin exon 1 are characterized by conformational heterogeneity that persists for wild type and pathogenic polyglutamine lengths. (<sup>§</sup>Co-first authors, <sup>\*</sup>Co-corresponding authors). *Journal of Molecular Biology*, **430**: 1442-1458.
  25. T.S. Harmon, A.S. Holehouse, **R.V. Pappu**. (2018). Differential solvation of intrinsically disordered linkers drives the formation of spatially organized droplets in ternary systems of linear multivalent proteins. *New Journal of Physics*, **20**: 045002.
  26. K. Garai, A.E. Posey, X. Li, E.M. Powers, J. Buxbaum, **R.V. Pappu**. (2018). Inhibition of amyloid beta fibril formation by monomeric human Transthyretin. *Protein Science*, **27**: 1252-1261.
  27. M.V. Staller, A.S. Holehouse, D. Swain-Lenz, R.K. Das, **R.V. Pappu**, B.A. Cohen. (2018). A high-throughput mutational scan of an intrinsically disordered acidic transactivation domain. *Cell Systems*, **6**: 444-455.e6.
  28. A.E. Posey, K.M. Ruff, T.S. Harmon, S.L. Crick, A. Li, M.I. Diamond, **R.V. Pappu**. (2018). Profilin reduces aggregation and phase separation of huntingtin N-terminal fragments by preferentially binding to soluble monomers and oligomers. *Journal of Biological Chemistry*, **293**: 3734-3746. (Article selected to be part of the JBC special issue on Amyloids, Prions, and Oligomers - <http://www.jbc.org/site/vi/amyloids/>)
  29. A.S. Holehouse, **R.V. Pappu**. (2018). Collapse transitions of proteins and the interplay amongst polypeptide backbone, sidechain, and solvent interactions. *Annual Review of Biophysics*, **47**: 19-39.
  30. A.S. Holehouse, **R.V. Pappu**. (2018). Functional implications of intracellular phase transitions. *Biochemistry*, **57**: 2415-2423.
  31. T. M. Franzmann, M. Jahnel, A. Pozniakovsky, J. Mahamid, A.S. Holehouse, E. Nüske, D. Richter, W. Baumeister, S.W. Grill, **R.V. Pappu**, A.A. Hyman, S. Alberti. (2018). Phase separation by a yeast prion protein promotes cellular fitness. *Science*, **359**: eaao5654.
  32. J. Fei<sup>\*</sup>, M. Jadalih<sup>§</sup>, T. S. Harmon<sup>§</sup>, I.T.S. Li, B. Hua, A. S. Holehouse, M. Reyer, Q. Sun, S. M. Freier, **R. V. Pappu**, K. V. Prasanth<sup>\*</sup>, T.J. Ha. Proteins and RNAs display multilayer organization in nuclear speckles. (2017). (<sup>\*</sup>co-corresponding authors; <sup>§</sup>equal contributors). *Journal of Cell Science*, **130**: 4180-4192.
  33. T.S. Harmon, A.S. Holehouse, M.K. Rosen, **R.V. Pappu**. (2017). Intrinsically disordered linkers determine the interplay between phase separation and gelation in multivalent proteins. *eLife*, **6**:30294. **In the news**: <https://source.wustl.edu/2017/12/engineers-uncover-design-principles-cellular-compartments>.
  34. A. S. Holehouse, **R.V. Pappu**. (2017). FUS zigzags its way to cross beta. *Cell*, **171**: 499-500.
  35. J. B. Warner IV<sup>§</sup>, K. M. Ruff<sup>§</sup>, P. S. Tan, E. A. Lemke, **R. V. Pappu**<sup>\*</sup>, H. A. Lashuel<sup>\*</sup>. Monomeric huntingtin exon 1 has similar overall structural features for wild type and pathological polyglutamine length. (2017). (<sup>§</sup>Co-first authors, <sup>\*</sup>Co-corresponding authors). *Journal of the American Chemical Society*, **139**: 14456 – 14469. **In the news**: <https://source.wustl.edu/2017/10/imaging-a-killer/>
  36. K. Sherry, R.K. Das, **R.V. Pappu**<sup>\*</sup>, D. Barrick<sup>\*</sup>. Control of transcriptional activity through de

- novo* design of sequence-to-conformation relationships of the intrinsically disordered RAM region of the Notch intracellular domain. (2017). (\*Co-corresponding authors). *Proceedings of the National Academy of Sciences USA*, **114**: E9243 – E9252. **Selected and evaluated by Faculty of 1000.**
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4. M.C. Cohan, A.M.P. Eddelbuettel, P.A. Levin, **R.V. Pappu**. Dissecting the functional contributions of the intrinsically disordered C-terminal tail of *B. subtilis* FtsZ. Submitted to the *Journal of Molecular Biology*.

### **Manuscripts in preparation**

1. J-M. Choi, F. Dar, A. Phillips, D. Mitrea, C.B. Stanley, R.W. Kriwacki, **R.V. Pappu**. Structure and dynamics of liquid-state condensates formed by multivalent proteins.
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9. M. J. Fossat, J-M. Choi, **R.V. Pappu**. Methods for multiscale simulations of intrinsically disordered proteins. *Invited review in preparation for Quarterly Reviews in Biophysics*.
10. M. J. Fossat, A.E. Posey, **R.V. Pappu**. *q*-Canonical Monte Carlo simulations help explain

the formation of single alpha helices by sequences with patterned clusters of oppositely charged residues. *In preparation for the Journal of the American Chemical Society.*

11. A.S. Holehouse<sup>†</sup>, **R.V. Pappu**<sup>†</sup>. PIMMS: A coarse-grained simulation engine for deriving sequence-encoded phase diagrams of intrinsically disordered proteins from large-scale simulations of phase separation. (<sup>†</sup>Co-corresponding authors).
12. A.S. Holehouse, K.M. Ruff, **R.V. Pappu**. Sequence encoding can enable spontaneous emulsification of condensates.
13. A.S. Holehouse<sup>§</sup>, **R.V. Pappu**<sup>§</sup>. Temperature sweep Metropolis Monte Carlo: A generalizable approach for enhanced sampling on rugged energy landscapes. *To be submitted to Journal of Chemical Physics.* (<sup>§</sup>Co-corresponding authors).
14. J. Lalmansingh, J.M. Choi<sup>†</sup>, **R.V. Pappu**<sup>†</sup>. Mapping of free energy basin landscapes enables comparisons of residue-specific intrinsic conformational preferences. (<sup>†</sup>Co-corresponding authors).
15. J.M. Lalmansingh, A.T. Keeley, K.M. Ruff, M.C. Cohan, **R.V. Pappu**, A.S. Holehouse<sup>†</sup>. CTraj: A general-purpose analysis library for analyzing atomistic simulations of intrinsically disordered proteins. *In preparation for eLife.*
16. A.E. Posey<sup>†</sup>, M. R. King, **R.V. Pappu**<sup>†</sup>. Experimental methods and theoretical analysis for mapping coexistence curves of biological macromolecules. (<sup>†</sup>Co-corresponding authors).
17. A.E. Posey<sup>†</sup>, T.S. Harmon, M. J. Fossat, **R.V. Pappu**<sup>†</sup>. Preferential neutralization of ionizable residues stabilizes alpha helices in sequences with patterned clusters of oppositely charged residues. (<sup>†</sup>Co-Corresponding authors).
18. K.M. Ruff<sup>†</sup>, **R.V. Pappu**<sup>†</sup>. Assessment of theoretical versus computational approaches for predicting the dimensions of intrinsically disordered proteins. (<sup>†</sup>Co-corresponding authors).
19. K. M. Ruff, A.E. Posey, F. Dar. **R.V. Pappu**. Polyphasic Linkage: A thermodynamic framework for understanding how ligand binding influences phase separation. *In preparation.*
20. K.M. Ruff, F. Dar, A.E. Posey, **R.V. Pappu**. Generalization of the polyphasic linkage formalism to model the impacts of networks of ligands on protein phase behavior.
21. K.M. Ruff<sup>†</sup>, M.O.G. Richardson, A.S. Holehouse, **R.V. Pappu**<sup>†</sup>. Evolutionary analysis of polar tracts and their sequence contexts across proteomes. (<sup>†</sup>Co-corresponding authors).
22. K.M. Ruff<sup>†</sup>, M.C. Cohan<sup>†</sup>, **R.V. Pappu**. Multiscale computational analysis of the impact of disordered tails on FtsZ conformations and assembly. (<sup>†</sup>Co-first authors)
23. X. Zeng, A. Chilkoti, **R.V. Pappu**. High-throughput computational design of intrinsically disordered protein polymers that undergo thermo-responsive phase behavior.
24. X. Zeng, **R.V. Pappu**. Combined theoretical and computational approach for calculating sequence-specific phase diagrams of intrinsically disordered protein polymers.
25. X. Zeng<sup>†</sup>, K.M. Ruff<sup>†</sup>, **R.V. Pappu**. Enhancing machine learning aided coarse-grained simulations of intrinsically disordered protein polymers using cycles of active learning and refinement. *In preparation for the Journal of Physical Chemistry B.* (<sup>†</sup>Co-first authors)
26. T.S. Harmon, S. Roberts, A. Chilkoti, **R.V. Pappu**. A physical model for programmable hysteresis in partially ordered protein polymers that undergo thermoresponsive phase transitions.

## **Software**

1. **PIMMS – Polymer Interactions in Multicomponent Mixtures:** PIMMS is a high performance lattice-based 2D/3D Monte Carlo simulation engine. It uses a bead-per-residue representation for intrinsically disordered proteins and unfolded RNA molecules (as well as synthetic polymers) and can be used to explore how polymer sequence architecture impacts phase transitions. It can also be deployed to explore the sequence-specific behavior of disordered systems such as intrinsically disordered proteins. It facilitates full phase space exploration as well as arbitrary sequence complexity and residue-residue parameterization. The parameters for PIMMS can be obtained purely via phenomenological approaches or through systematic machine learning aided protocols developed in the Pappu lab. The primary developer and distributor of PIMMS is Dr. Alex S. Holehouse, currently a postdoctoral scientist in the Pappu lab and soon to be an independent PI in the Department of Biochemistry & Molecular Biophysics at Washington University's School of Medicine. Updates regarding PIMMS, which is designed to be an open-source software package are available at <http://pimms.xyz>.
2. **LASSI – Lattice simulations of Sticker and Spacer Interactions:** This open-source lattice-based Monte Carlo engine is designed to enable the simulation of coarse-grained descriptions of linear and branched multivalent proteins comprising of intrinsically disordered regions and folded domains of proteins. Based loosely on the bond fluctuation model, LASSI enables the incorporation of directional interactions and allows for coarse-grained models of different resolutions that are parameterized either using phenomenology or through systematic learning approaches based on transfer of information from atomistic simulations. LASSI enables the simulation of phase behavior, including the extraction of full phase diagrams based on a range of order parameters, for multi-component systems. It can be used as a predictive tool, a tool to enable design of bespoke phase behavior, or as a complement to experimental approaches where it can be deployed to explain measured phase behavior. The primary developers of LASSI are Furqan Dar, PhD student in the Pappu lab, and Dr. Jeong-Mo Choi, postdoctoral alumnus of the Pappu lab.
3. **CIDER and localCIDER – Classification of Intrinsically Disordered Ensemble Regions:** CIDER is a webserver being developed by the Pappu Lab for calculating parameters relating to disordered protein sequences. Specifically, this will calculate various parameters that help translate primary sequence information into a better understanding of the conformational properties of disordered proteins. The server provides a ready-made annotation on our diagram-of-states for IDPs. In addition, predictions from CIDER yield insights regarding emergent properties of IDPs such as the conversion of polyampholytic IDPs to polyelectrolytes and vice versa as the result of phosphorylation. Alex S. Holehouse is the main developer of CIDER. Available for free download from <http://pappulab.github.io/localCIDER/>.
4. **CAMPARI:** A simulation engine that integrates several sampling methodologies including molecular dynamics, rigid body dynamics, Langevin dynamics, novel torsional molecular dynamics, and an extensive code base for Monte Carlo sampling of biological macromolecules (mainly proteins, but also supports nucleic acids). The package was developed mainly by Andreas Vitalis, building on a skeleton that was developed by Pappu in the early days of the lab's existence. CAMPARI can be used for efficient calculation of free energies of solvation, neat liquids, complex fluids, and peptides in explicit solvent. However, the main advantage is its support of implicit solvent paradigms for use in Monte Carlo simulations. It is this aspect that makes CAMPARI unique. Significant work in the Pappu lab has shown that Monte Carlo simulations are extremely useful for efficient sampling of conformational space in conjunction with implicit solvation

models such as ABSINTH, which was also developed by Andreas Vitalis in the Pappu Lab. This model may be viewed as an efficient interpolation between the EEF1 and generalized Born (GB) paradigms. CAMPARI will also integrate support for the major variants of the GB framework to facilitate comparative calculations between different implicit solvation paradigms. The CAMPARI engine, first built around the ABSINTH model, was designed for answering questions regarding conformational and binding equilibria of intrinsically disordered polypeptides. However, it has wider usage and has gained acceptance as a *bona fide* engine in the biomolecular simulation community. CAMPARI is available for free download from <http://campari.sourceforge.net>.

5. **NPO – a nanoparticle ontology:** Data generated from cancer nanotechnology research are so diverse and large in volume that it is difficult to share and efficiently use them without informatics tools. In particular, ontologies that provide a unifying knowledge framework for annotating the data are required to facilitate the semantic integration, knowledge-based searching, unambiguous interpretation, mining and inferencing of the data using informatics methods. We developed a NanoParticle Ontology (NPO) within the framework of the Basic Formal Ontology (BFO), and implemented in the Ontology Web Language (OWL) using well-defined ontology design principles. The NPO is accessible online at <http://www.nano-ontology.org/>. Dr. Dennis Thomas developed this effort, sponsored by the National Cancer Institute.

## **Grants**

### **CURRENT**

#### **National Science Foundation, DMR 1729783 – DMREF Collaborative Research**

**Title:** *High throughput exploration of sequence space of peptide polymers that exhibit aqueous demixing phase behavior*

**Funding Period:** October 01, 2017 – September 30, 2021

**Principal Investigator:** Ashutosh Chilkoti, Duke University

**Role in the project:** *Co-Principal Investigator* with Stefan Zauscher (Duke University)

#### **Human Frontier Science Program (HFSP), RGP0034/2017**

**Title:** *Elucidating the molecular logic of membrane-free compartment function and assembly*

**Funding period:** September 01, 2017 – August 31, 2020

**Role in the project:** *Co-Principal Investigator* with Simon Alberti (Max Planck Institute, Dresden) and Stephen W. Michnick (University of Montreal)

#### **St. Jude Research Collaborative, Sponsored by St. Jude Children's Research Hospital,**

**Title:** *Biology and Biophysics of Membraneless Organelles*

**Funding period:** February 01, 2017 – January 31, 2022

**Role in the project:** *Project Principal Investigator*, Computational & Theoretical Modeling

#### **National Science Foundation, MCB-1614766**

**Title:** *Multiscale Modeling of Phase Transitions Driven by Multivalency and Disordered Proteins*

**Funding period:** July 15, 2016 – July 14, 2020

**Role in the project:** *Principal Investigator*

#### **National Institutes of Health (NINDS), 5R01NS056114-10-14**

**Title:** *Role of chain length and sequence contexts on polyglutamine oligomerization*

**Funding period:** July 01, 2016 – June 30, 2021

**Role in the project:** *Principal Investigator*

**National Institutes of Health, 1R01NS089932-A1**

**Title:** *Mechanism of modulation of huntingtin exon 1 aggregation by profilin*

**Funding period:** April 01, 2016 – December 31, 2020

**Principal Investigators:** Rohit Pappu (contact PI), Marc I. Diamond, and Ralf Langen

**Role in the project:** *Main Principal Investigator* in Multi-PI grant

**PENDING**

**National Institutes of Health, 1R01 GM131148**

**Title:** *Sequence Determinants of the conformations and functions of disordered proteins that undergo multisite phosphorylation*

**Principal Investigators:** Tanja Mittag (contact PI), Rohit Pappu (co-PI)

**Co-Investigators:** Barak Cohen, Benjamin Schuler

**Role in the project:** *Principal Investigator* in Multi-PI grant

**National Institutes of Health, 1RM1 GM136509**

**Title:** *Multiscale investigations of stress granule assembly and dynamics*

**Principal Investigators:** Rohit Pappu (contact PI), Tanja Mittag (co-PI), Andrea Soranno (co-PI), J. Paul Taylor (co-PI);

**Role in the project:** *Co-Principal Investigator* in collaborative grant

**COMPLETED**

**National Institutes of Health, 1R01GM108785**

**Title:** *Signal transduction by ERBB2 / ERBB3 oligomers*

**Funding period:** October 01, 2013 – March 31, 2017

**Principal Investigator:** Linda J. Pike

**Role in the project:** *Co-Investigator*

**National Institutes of Health (NINDS), 5R01NS056114-05-09**

**Title:** *Role of chain length and sequence contexts on polyglutamine oligomerization*

**Funding period:** July 01, 2011 – June 30, 2016

**Role in the project:** PI

**National Science Foundation, MCB-0718924**

**Title:** *Phase behavior of intrinsically disordered proteins*

**Funding period:** September 01, 2011 – August 31, 2015

**Role in the project:** Principal Investigator

**National Institutes of Health, 1S10OD018091 Shared Instrumentation Grant**

**Title:** *GPU computing resource to enable innovation in imaging and network biology*

**Funding period:** April 01, 2014 – March 31, 2015

**Role in the project:** Co-PI with Fred Prior

**National Institutes of Health (NINDS), 5R01NS056114**

**Title:** *Atomistic studies of nucleation and oligomerization in polyglutamine aggregation*

**Funding period:** April 15, 2007 – June 30, 2012

**Role in the project:** PI

**National Science Foundation, MCB-0718924**

**Title:** *Conformational equilibria of intrinsically disordered proteins*

**Funding period:** September 01, 2007 – August 31, 2011

**Role in the project:** PI

**Hope Center for Neurological Disorders, Washington University School of Medicine**

Translational Neuroscience Pilot Project

**Title:** *Mechanism of Huntingtin Aggregation Regulated by Profilin*

**Funding period:** June 01, 2011 – May 31, 2013

**Role in the project:** Principal Investigator

**Co-Investigator:** Marc Diamond

**National Institutes of Health (NCI) U54 CA-119342**

**Title:** *An informatics resource for targeted nanoparticle therapeutics*

NCI Center grant to the Siteman Center for Cancer Nanotechnology Excellence (Center grant

**PI:** Samuel Wickline)

**Funding period:** February 01, 2006 – January 31, 2011

**Role in the project:** Project Principal Investigator

**Project Co-Investigators:** David Sept, Nathan Baker

**Pfizer Inc., St. Louis, MO**

**Title:** *Modeling the aggregation of therapeutic monoclonal antibodies*

**Funding period:** October 31, 2009 – September 30, 2010

**Role in the project:** Project Principal Investigator

**National Institutes of Health (NCI) Integrated Cancer Research Workspace 94358NBS23**

*Subcontract for with Booz Allen Hamilton for Nanotechnology working group*

**Funding period:** April 01, 2008 – February 14, 2009

**Role in the project:** Principal Investigator

**National Science Foundation, MCB-0416766**

**Title:** *Studying the origin of conformational preferences in unfolded proteins*

**Funding period:** September 01, 2004 – August 31, 2007

**Role in the project:** Principal Investigator

**Hope Center for Neurological Disorders, Washington University School of Medicine**

Translational Neuroscience Pilot Project

**Title:** *Investigation of structural changes induced in amyloid A $\beta$  fibrils by polyphenols*

**Funding period:** January 01, 2007 – December 31, 2007

**Role in the project:** Co-Investigator (PI: Jin-Moo Lee)

**Fidelity Foundation**

**Title:** *Studies on the process of aggregation in Huntington's disease*

**Funding period:** January 01, 2006 – December 31, 2006

**Role in the project:** Principal Investigator

**March of Dimes Birth Defects Foundation, Basil O'Connor Starter Scholar Award**

**Title:** *Factors that determine amyloid formation in polyglutamine disorders*

**Funding period:** February 01, 2004 – June 30, 2006

**Role in the project:** Principal Investigator

**National Institutes of Health, NIA P50 AG05681-20**

Pilot grant from Alzheimer's Disease Research Center, Washington University

**Title:** *Toward a molecular understanding of polyglutamine disorders*

**Funding period:** May 01, 2003 – April 30, 2004

**Role in the project:** Principal Investigator

**RESEARCH MENTORING**

***Current Postdoctoral and Research Scientists***

1. **Martin Fossat**, Postdoctoral Scientist
2. **Alex S. Holehouse**, Postdoctoral Scientist
3. **Matthew R. King**, Postdoctoral Scientist
4. **Ammon E. Posey**, Research Scientist
5. **Kiersten M. Ruff**, Research Scientist
6. **Max V. Staller**, Co-mentored Postdoctoral Scientist (primary mentor: Barak A. Cohen)
7. **Xiangze Zeng**, Postdoctoral Scientist

***Current Doctoral Students (alphabetical order)***

1. **Megan C. Cohan**, Doctoral Student, Department of Biomedical Engineering. Projected graduation date, August 2020.
2. **Furqan Dar**, Doctoral Student, Department of Physics, Projected graduation date, December 2021.
3. **Mina Farag**, MD / PhD Student, Department of Biomedical Engineering, Projected graduation date, Spring 2023.
4. **Jared M. Lalmansingh**, Doctoral Student, Department of Physics. Projected graduation date, December 2020.
5. **Andrew Z. Lin**, Doctoral Student, Division of Biology & Biomedical Sciences, Program Plant and Microbiology. Projected graduation date, August 2021.

***Current Undergraduate Students (alphabetical order)***

1. **Anna M. P. Eddebuettel**, Third-year student in the Department of Biomedical Engineering. Scheduled to graduate, May 2020.

***Current Research Staff (alphabetical order)***

1. **Catherine Kornacki**, Laboratory Research Technician, August 2019 onward.

***Lab alumni – reverse chronological order***

1. **Garrett Ginell**, Junior Computational Research Scientist, August 2018 – August 2019
2. **Jeong-Mo Choi**, Postdoctoral Scientist, August 2016 – April 2019.
3. **Mary O. G. Richardson**, Junior Research Scientist and Scientific Programmer, August 2017 – June 2018.
4. **Sang Eun Jee**, Postdoctoral Scientist, November 2016 – October 2017.
5. **Kourtney L. Kroll**, Undergraduate researcher, Summer 2017.
6. **Divya Natarajan**, MD Student, Washington University School of Medicine, graduation date, May 2020.

7. **Tyler S. Harmon**, Doctoral Student, Department of Physics. Received PhD in April 2017. Currently, postdoctoral scientist at the Max Planck Institute in Dresden, Germany.
8. **Rajni Verma**, Postdoctoral Scientist, October 2015 – June 2016.
9. **Rahul K. Das**, Postdoctoral Scientist, September 2009 – August 2015.
10. **Anuradha Mittal**, Postdoctoral Scientist. Currently, a senior Bioinformatics Scientist at Affymetrix, San Francisco, CA.
11. **Laruen M. Bedell**, Undergraduate Student, Department of Biomedical Engineering.
12. **Kanchan Garai**, Research Assistant Professor and Research Scholar, Center for Biological Systems Engineering. Currently, Assistant Professor at the TIFR Centre for Interdisciplinary Studies, Hyderabad, India.
13. **James Ahad**, Undergraduate student, Biomedical Engineering. Currently, an MSTP student at Case Western University School of Medicine.
14. **Scott L. Crick**, Doctoral Student, Biomedical Engineering. Received Ph.D. in August 2011. Completed a two-year Postdoc in the lab in July 2013. Currently working as a Licensing Associate in the Office of Technology Management at Washington University in St. Louis.
15. **Siddique J. Khan**, Postdoc, April 2012 – May 2013.
16. **Nicholas Lyle**, Doctoral Student, Computational & Systems Biology Program, Division of Biology & Biomedical Sciences. Received Ph.D. in May 2013. Currently, Senior Scientific Analyst at Partek Inc., Chesterfield, MO.
17. **Tony Wang**, Summer undergraduate researcher, Summer 2012.
18. **Marta Wells**, Summer undergraduate research, Summer 2011 and 2012. Currently a graduate student in Computational Biology at Carnegie-Mellon University.
19. **Albert H. Mao**, M.D.-Ph.D. student, Computational & Molecular Biophysics Program, Division of Biology & Biomedical Sciences. Received Ph.D. in August 2012. Currently doing his residency in anesthesiology at Harvard Medical School, Beth Israel Hospital.
20. **Jordan Nick**, Summer HHMI undergraduate fellow, Summer 2011.
21. **Kelly Culhane**, SURF student, Summer 2011. Currently, a Ph.D. student in Biochemistry & Biophysics at Yale University.
22. **Aditya Radhakrishnan**, Master's student (BS/MS), Summer 2010 – Summer 2011. Currently Ph.D. student in Computational & Molecular Biophysics at Johns Hopkins University.
23. **Nil Gural**, Undergraduate researcher, Spring 2011. Currently Ph.D. student in the Harvard-MIT Health Sciences Program.
24. **Adam T. Steffen**, Scientific programmer, Summer 2007 – December 2010. Currently a software engineer at Partek Inc., Chesterfield, MO.
25. **Alexander French**, Undergraduate researcher, Summer 2010. Currently graduate student in Biochemistry & Biophysics at the University of Chicago.
26. **Anil Kumar**, Postdoctoral scientist, December 2009 – August 2010. Currently a postdoc at the University of Toronto in synthetic organic chemistry.

27. **Caitlin L. Chicoine**, Undergraduate researcher, Summer of 2009, Graduated from Washington University, School of Engineering & Applied Sciences Valedictorian.
28. **Matthew A. Wyczalkowski**, Biomedical Engineering. Received his Ph.D. in December 2009. Currently a senior postdoctoral scientist at The Genome Institute, Washington University School of Medicine.
29. **Tim E. Williamson**, Molecular Biophysics Program, Division of Biology & Biomedical Sciences and Staff Scientist. Received MS in May 2009. Currently working as a technical officer at Monsanto Inc., St. Louis, MO.
30. **Andreas Vitalis**, Doctoral Student, Molecular Biophysics Program, Division of Biology & Biomedical Sciences. Received his Ph.D. in June 2009. Currently an independent research scientist completing his research habilitation in the Department of Biochemistry at the University of Zurich.
31. **Alan A. Chen**, Doctoral Student, Molecular Biophysics Program, Division of Biology & Biomedical Sciences. Received his Ph.D. in May 2009. Currently assistant professor of chemistry the University of Albany, RNA Institute.
32. **Jose Pulido**, Undergraduate Trainee, May 2008 – October 2008.
33. **Xiaoling Wang**, Research scientist, September 2004 – February 2008.
34. **Hoang T. Tran**, Doctoral student, Biomedical Engineering. Received Ph.D. in December 2007. Currently a staff scientist in the drug discovery at the M.D. Anderson Cancer Center in Houston, TX.
35. **Alexander N. Drozdov**, Postdoctoral Scientist, September 2002 – April 2004.
36. **Magdalena Fus**, Undergraduate Trainee, September 2004 – May 2005.
37. **Tirath Patel**, Undergraduate Trainee, September 2004 – May 2005.

#### ***Awards & Honors to Members of the Lab – Reverse Chronological Order***

- Ms. Megan C. Cohan, graduate student in the Pappu lab, was elected to be the graduate student representative to the council of the Intrinsically Disordered Proteins subgroup of the Biophysical Society for the period between April 01, 2019 and March 31, 2020.
- Dr. Jeong-Mo Choi, postdoctoral fellow in the Pappu lab, received a travel award to attend and present his work at the 63<sup>rd</sup> Annual Meeting of the Biophysical Society, Baltimore, March 2019.
- Ms. Megan C. Cohan, graduate student in the Pappu lab, was elected to the board of BALSAs for the 2019-2020 period. This is the Biotechnology and Life Sciences Advising Group, which is a non-profit organization “that provides consulting services for a variety of clients, ranging from individual faculty members up through multinational corporations”.
- Mr. Jared Lalmansingh, physics PhD student in the Pappu lab, received Washington University’s university’s 2019 Gerry and Bob Virgil Ethic of Service Award for his contributions to the Young Scientist’s Program, an outreach effort to draw students from the inner cities into science.
- Dr. Alex S. Holehouse, postdoc in the Pappu lab, received the 2019 MoISSI “seed” software fellowship from the Molecular Sciences and Software Institute to develop a computational pipeline for modeling the evolution and sequence-ensemble relationships of intrinsically disordered proteins.

- Dr. Max Staller, joint postdoc in the Cohen and Pappu labs, was chosen to give two lectures as part of the Gene Regulatory Networks in Development course at the Molecular Biology Laboratory in Woods Hole, MA in October 2018.
- Mr. Conor O'Neill, first year Biomedical Engineering graduate student in the Pappu lab, was matched with the Kent and Bonnie Lattig fellowship in the Center for Biological Systems Engineering at Washington University in St. Louis for the 2018-2019 academic year.
- Dr. Alex S. Holehouse, postdoctoral fellow in the Pappu lab, received an award for one of the three best posters presented by postdocs at the 2018 Gordon Research Conference on Intrinsically Disordered Proteins held in Les Diablerets, Switzerland.
- Ms. Megan C. Cohan, graduate student in the Pappu lab, was elected by her peers to co-chair the 2020 Gordon Research Seminar that will precede the 2020 Gordon Research Conference on Intrinsically Disordered Proteins.
- Dr. Kiersten M. Ruff, postdoctoral fellow in the Pappu lab, was chosen to give a special talk at the 2018 Gordon Research Conference on Intrinsically Disordered Proteins held in Les Diablerets, Switzerland.
- Ms. Megan C. Cohan, graduate student in the Pappu lab, was chosen to speak at the 2018 Gordon Research Seminar (GRS) that preceded the 2018 Gordon Research Conference on Intrinsically Disordered Proteins in Les Diablerets, Switzerland, July 2018.
- Ms. Megan C. Cohan, graduate student in the Pappu lab, was chosen to receive a travel award, based on her submitted abstract, to present a poster at the EMBO | EMBL Symposium on Cellular Mechanisms Driven by Liquid Phase Separation, May, 2018.
- Dr. Kiersten M. Ruff, postdoctoral fellow in the Pappu lab, was chosen, based on her submitted abstract, to speak at the EMBO | EMBL Symposium on Cellular Mechanisms Driven by Liquid Phase Separation, May, 2018.
- Ms. Mary O.G. Richardson, scientific programmer and junior research scientist in the Pappu lab received the prestigious NSF graduate student fellowship for her PhD studies in Computational Biology.
- Dr. Jeong-Mo Choi, postdoctoral fellow in the Pappu lab, was chosen to speak in a platform session on intrinsically disordered proteins at the 62<sup>nd</sup> Annual Meeting of the Biophysical Society, San Francisco, February 2018.
- Dr. Kiersten M. Ruff, postdoctoral fellow in the Pappu lab, was chosen to speak in a platform session on intrinsically disordered proteins at the 62<sup>nd</sup> Annual Meeting of the Biophysical Society, San Francisco, February 2018.
- Dr. Jeong-Mo Choi, postdoctoral fellow in the Pappu lab, was chosen to give the Molecular Kinetics sponsored postdoctoral talk at the 2018 symposium of the Intrinsically Disordered Proteins subgroup to be held in conjunction with the 62<sup>nd</sup> Annual Meeting of the Biophysical Society, San Francisco, February 2018.
- Dr. Alex S. Holehouse, postdoctoral fellow in the Pappu lab, was one of the invited speakers in the symposium organized by the Biopolymers In Vivo Subgroup at 62<sup>nd</sup> Annual Meeting of the Biophysical Society, San Francisco, February 2018.
- Dr. Alex S. Holehouse, postdoctoral fellow in the Pappu lab, was chosen to give a short talk at the 2018 Protein Folding Dynamics Gordon Research Conference, Galveston, TX, January 2018.
- Ms. Megan C. Cohan, Biomedical Engineering Graduate Student in the Pappu Lab, received a travel award to attend the 2017 annual meeting of the Biomedical Engineering Society to be held in Phoenix, AZ, October 2017.

- Dr. Kiersten M. Ruff, postdoctoral fellow in the Pappu Lab, was chosen to give a short talk at the 2017 FASEB Science Research Conference on Protein Aggregation in Health and Disease in Steam Boat Springs, CO, June 2017.
- Ms. Megan C. Cohan, Biomedical Engineering Graduate Student in the Pappu Lab, was chosen to give a student talk at the annual meeting of the NSF sponsored Protein Folding Consortium in Berkeley, CA, June 2017.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab gave a talk at the weeklong workshop on the Physics of Cellular Adaptation held at the Bellairs Institute in Barbados, April 2017.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab, Recipient of the Spencer T. and Ann W. Olin for superior accomplishments in biomedical research doctoral students at Washington University, February 2017.
- Mr. Tyler S. Harmon, Physics Graduate Student in the Pappu Lab Recipient of the SRAA Award for Best Poster, 61<sup>st</sup> Annual Meeting of the Biophysical Society, Award Sponsored by the Intrinsically Disordered Proteins Subgroup, February 2017.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab gave an invited platform session talk and chaired this session at the 61<sup>st</sup> Annual Meeting of the Biophysical Society, February 2017.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab, received a student speaker award at the 23<sup>rd</sup> Annual retreat of the DBBS Computational & Molecular Biophysics Graduate Program for his talk entitled *Phase separation of intrinsically disordered proteins yields low-density "empty" liquids*.
- Ms. Megan C. Cohan, Biomedical Engineering Graduate Student in the Pappu Lab, received a travel award to attend the fourth biennial Intrinsically Disordered Proteins Gordon Research Conference held in Les Diablerets, Switzerland in June 2016.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab, received a travel award to attend the fourth biennial Intrinsically Disordered Proteins Gordon Research Conference held in Les Diablerets, Switzerland in June 2016.
- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Student in the Pappu Lab, was one of the awardees for best posters presented at the fourth biennial Intrinsically Disordered Proteins Gordon Research Conference held in Les Diablerets, Switzerland in June 2016.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab was elected as co-chair of the 2018 Graduate Research Seminar that is to accompany the fifth biennial Intrinsically Disordered Proteins Gordon Research Conference.
- Mr. Tyler S. Harmon, Physics Graduate Student in the Pappu Lab was named as one of the CBSE Graduate Student Scholar by the Center for Biological Systems Engineering at Washington University for the period between July 01, 2016 and June 30, 2017.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab was named as the Kent and Bonnie Lattig CBSE Graduate Student Scholar by the Center for Biological Systems Engineering at Washington University for the period between July 01, 2016 and June 30, 2017.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab selected to give an invited talk at the Gordon Research Seminar preceding the Gordon Research Conference on Intrinsically Disordered Proteins, Les Diablerets, Switzerland, June 2016.

- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab gave an invited platform session talk at the 60<sup>th</sup> Annual Meeting of the Biophysical Society, February 2016.
- Mr. Tyler S. Harmon, Physics Graduate Student in the Pappu Lab gave an invited platform session talk at the 60<sup>th</sup> Annual Meeting of the Biophysical Society, February 2016.
- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Student in the Pappu Lab, was named the first Kent and Bonnie Lattig CBSE Graduate Student Scholar by the Center for Biological Systems Engineering at Washington University in St. Louis for the period between July 2015 and June 2016.
- Mr. Tyler S. Harmon, Physics Graduate Student in the Pappu Lab, was chosen to give the opening talk at the 2015 edition of the annual meeting of the NSF sponsored Protein Folding Consortium, Berkeley, May 2015.
- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Student in the Pappu Lab, was reappointed as a CBSE Graduate Student Scholar by the Center for Biological Systems Engineering at Washington University in St. Louis for the period between July 2015 and June 2016.
- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Students in the Pappu Lab was chosen to speak at the Princeton Workshop on Intracellular Phase Transitions, April 2015.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab, Recipient of the SRAA Award for Best Poster, 59<sup>th</sup> Annual Meeting of the Biophysical Society, Award Sponsored by the Intrinsically Disordered Proteins Subgroup, February 2015.
- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Student in the Pappu Lab, Selected to deliver a platform session talk at the 59<sup>th</sup> annual meeting of the Biophysical Society, Baltimore, MD, February 2015.
- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Student in the Pappu Lab, was appointed as a CBSE Graduate Student Scholar by the Center for Biological Systems Engineering at Washington University in St. Louis for the period between July 2014 and June 2015.
- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Student in the Pappu Lab, Selected to speak at the first Graduate Research Seminar preceding the 3<sup>rd</sup> Gordon Research Conference on Intrinsically Disordered Proteins, Stonehill College, MA, June 2014.
- Mr. Albert H. Mao, MSTP and Computational & Molecular Biophysics Graduate Student in the Pappu Lab, received Washington University's Spencer T. Olin prize for outstanding graduate research, April 2014.
- Mr. Alex S. Holehouse, Computational & Molecular Biophysics Graduate Student in the Pappu Lab, Recipient of the SRAA Award for Best Poster, 58<sup>th</sup> Annual Meeting of the Biophysical Society, Award Sponsored by the Intrinsically Disordered Proteins Subgroup, February 2014.
- Dr. Anuradha Mittal, Postdoc in the Pappu Lab, Selected to give a platform session talk at the 58<sup>th</sup> Annual Meeting of the Biophysical Society, San Francisco, February 2014.
- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Student in the Pappu Lab, Recipient of the SRAA Award for Best Poster, 58<sup>th</sup> Annual Meeting of the Biophysical Society, Award Sponsored by the Intrinsically Disordered Proteins Subgroup, February 2014.

- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Student in the Pappu Lab, Received Honorable Mention from National Science Foundation in response to Graduate Student Fellowship, with Enhanced Access to Cyberinfrastructure Resources, Including XSEDE Supercomputing Time to Support Research Toward Completion of Graduate Program, 2013.
- Ms. Kiersten M. Ruff, Computational & Systems Biology Graduate Student in the Pappu Lab, Recipient of the SRAA Award for Best Poster, 57<sup>th</sup> Annual Meeting of the Biophysical Society, Award Sponsored by the Intrinsically Disordered Proteins Subgroup, 2013.
- Dr. Scott L. Crick, Recipient of the National Institutes of Health, National Research Service Award for Postdoctoral Research following the completion of thesis work and a two-year postdoctoral stint in the Pappu Lab, 2013.
- Mr. Nicholas Lyle, Computational & Systems Biology Graduate Student, Best Poster Award, Intrinsically Disordered Proteins, Gordon Research Conference, 2012.
- Dr. Scott L. Crick, Postdoc in the Pappu Lab, Selected to deliver a Platform Session Talk at the 56<sup>th</sup> Annual Meeting of the Biophysical Society, San Diego, CA, 2012.
- Dr. Rahul K. Das, Postdoc in the Pappu Lab, Recipient of the Molecular Kinetics Postdoctoral Speaking Award and Honorarium, 5<sup>th</sup> Annual Symposium of the Intrinsically Disordered Proteins Subgroup at the 55<sup>th</sup> Annual Meeting of the Biophysical Society, Baltimore, MD, 2011.
- Mr. Albert H. Mao, MSTP and Computational & Molecular Biophysics Graduate Student in the Pappu Lab, Recipient of the SRAA Award for Best Poster, 55<sup>th</sup> Annual Meeting of the Biophysical Society, Baltimore, MD, Award Sponsored by the Intrinsically Disordered Proteins Subgroup, 2011.
- Mr. Scott L. Crick, Biomedical Engineering Graduate Student in the Pappu Lab, Selected to deliver a Platform Symposium Talk at the 54<sup>th</sup> Annual Meeting of the Biophysical Society, San Francisco, CA, 2010.
- Dr. Matthew A. Wyczalkowski, Former Biomedical Engineering Graduate Student, Recipient of the National Institutes of Health, National Research Service Award for Postdoctoral Research following the completion of thesis work in the Pappu Lab, 2010.
- Dr. Alan A. Chen, Former Computational & Molecular Biophysics Graduate Student, Recipient of the National Institutes of Health, National Research Service Award for Postdoctoral Research following the completion of thesis work in the Pappu Lab, 2010.
- Dr. Andreas Vitalis, Computational & Molecular Biophysics Graduate Student Empiris Award for Research in Brain Diseases, Award received for Ph.D. thesis work performed in the Pappu Lab, 2010.

***Other Invited talks: Fall 2001 – present (reverse chronological order)***

1. Princeton University, Department of Chemical & Biochemical Engineering, May 2018
2. University of California, Berkeley, Molecular and Cell Biology, April 2017
3. Hospital for Sick Kids, Toronto, Molecular Medicine Seminar Series, March 2017
4. Washington University, Department of Genetics Seminar Series, February 2017
5. McGill University, Department of Biology, January 2017
6. Princeton University, Department of Chemical & Biochemical Engineering, December 2016
7. Tata Institute for Fundamental Research, Mumbai, India, November 2016
8. Department of Biomedical Engineering, Washington University in St. Louis, October 2016

9. MRC Laboratory of Molecular Biology, Cambridge, UK, June 2016
10. Johns Hopkins University, Department of Materials Science, April 2016
11. Hiroshima University, December 2015
12. Duke University, NSF-MRSEC, April 2015
13. University of British Columbia, Center for High-Throughput Biology, April 2015
14. Washington University in St. Louis, Biophysical Evening, January 2015
15. University of Texas Southwestern Medical Center, Department of Biophysics, December 2014
16. University of Cambridge, Department of Chemistry, November 2014
17. MRC Laboratory of Molecular Biology, Cambridge, UK, November 2014
18. Stony Brook University, Laufer Center for Quantitative Biology, November 2014
19. Kansas State University, Department of Biochemistry, April 2014
20. Ohio State University, Department of Biochemistry, Biophysics Program, April 2014
21. University of Montana, Department of Chemistry & Biochemistry, February 2014
22. Johns Hopkins University, Department of Biophysics, December 2013
23. University of Wisconsin-Madison, Department of Chemical & Biochemical Engineering, December 2013
24. University of Zurich, Department of Biochemistry, September 2013
25. The Scripps Research Institute, Department of Molecular & Experimental Medicine, July 2013
26. University of Texas Southwestern Medical Center, University Colloquium, July 2013
27. University of Texas Southwestern Medical Center, Green Center for Systems Biology, Special Seminar, July 2013
28. Symposium on Physics and Biology of Strongly Fluctuating Proteins, University of Maryland, May 2013
29. American Chemical Society Meeting, New Orleans, April 2013
30. University of Chicago, Institute for Biophysical Dynamics, April 2013
31. March Meeting of the American Physical Society, Baltimore, March 2013
32. CUNY City College of New York, Structural Biology Colloquium, March 2013
33. 57<sup>th</sup> Annual Meeting of the Biophysical Society, Philadelphia, Protein Electrostatics Symposium
34. University of Minnesota, Department of Biomedical Engineering, December 2012
35. Protein Folding Consortium, Pls meeting, Chicago, November 2012
36. International Symposium on Protein Folding, National Centre for Biological Sciences, Bangalore, India, October 2012
37. Protein Folding Consortium, SUNY Stony Brook, NY, June 2012
38. Biopolymers Gordon Research Conference, Newport, RI, June 2012
39. Hope Center for Neurological Disorders, Washington University, May 2012
40. Arizona State University, Center for Biological Physics, Tempe, AZ, April 2012
41. University of South Florida, Department of Physics, Tampa, FL, March 2012
42. St. Jude Children's Research Hospital, Memphis, TN, January 2012
43. CCP 2011 – Conference on Computational Physics, Knoxville, TN, November 2011
44. National Science Foundation Workshop on Future of RNA and Protein Folding, Arlington, VA, September 2011
45. University of Leeds, UK, July 2011

46. MRC Laboratory of Molecular Biology, Cambridge, UK, July 2011
47. Annual Meeting of the Protein Folding Consortium, UC Berkeley, June 2011
48. University of Texas Southwestern Medical School, May 2011
49. The Scripps Research Institute, Molecular & Experimental Medicine, May 2011
50. Washington University School of Medicine, Computational & Molecular Biophysics Program, Biophysical Evening Series, May 2011
51. Midwest Conference on Protein Folding, Assembly, and Molecular Motions, May 2011
52. 55<sup>th</sup> Annual Meeting of the Biophysical Society, Baltimore, MD, March 2011
53. Department of Biochemistry, Cornell Weill Medical College, December 2010
54. ACS Midwest Regional Meeting Symposium on Protein Folding, October, 2010
55. Intrinsically disordered proteins, Keynote Session Chair, Gordon Research Conference, North Carolina, July 2010.
56. Protein folding pathways workshop, Arizona State University, May 2010.
57. University of Pittsburgh School of Medicine, March 2010.
58. Yale University, Biophysics program, January 2010.
59. Protein folding and dynamics, Gordon Research Conference, January 2010.
60. The Scripps Research Institute, Molecular & Experimental Medicine, November 2009.
61. 23<sup>rd</sup> Annual Gibbs conference on Biothermodynamics, Carbondale, IL, October 2009.
62. Tata Institute of Fundamental Research, Mumbai, India, August 2009.
63. Argonne National Laboratory, Biology Division, August 2009.
64. Northwestern University, Biochemistry, Molecular Biology & Cell Biology, July 2009.
65. Telluride Scientific Research Conference, Workshop on RNA dynamics, July 2009.
66. FASEB Amyloid meeting, Snowmass, CO, June-July 2009.
67. University of California Berkeley, Department of Bioengineering, April 2009.
68. MIT-Whitehead Institute, Department of Biology, March 2009.
69. Annual Meeting of the Biophysical Society, Invited Workshop, Boston, MA, March 2009.
70. Department of Chemistry & Biochemistry, University of Massachusetts, Amherst, MA, February 2009
71. Rice University, Department of Chemistry, November 2009.
72. Rensselaer Polytechnic Institute, Biocomputation seminar series, October 2008.
73. 22<sup>nd</sup> Annual Symposium of the Protein Society, July 2008, San Diego, CA
74. Protein Electrostatics Workshop, Telluride, CO, July 2008
75. Biopolymers Gordon Research Conference, Newport, RI, June 2008.
76. University of Texas Austin, Department of Biomedical Engineering, April 2008.
77. University of Oregon, Department of Chemistry, April 2008.
78. Intrinsically Disordered Proteins subgroup meeting, Annual meeting of the Biophysical Society, Long Beach, CA, February 2008
79. The Scripps Research Institute, Department of Chemistry, La Jolla, CA, September 2007.
80. Gordon Research Conference: Proteins, June 2007, Holderness, New Hampshire.
81. Washington University in St. Louis, Biophysical Evening Series, December 2007.
82. University of California in Santa Barbara, Department of Chemistry, April 2007.
83. University of Delaware, Department of Chemistry and Biochemistry, April 2007.
84. Indiana University, Computational Biology and Bioinformatics, September 2006.
85. National Cancer Institute, Frederick, August 2006.

86. FASEB Amyloid Meeting, Snowmass, Colorado, June 2006.
87. DIMACS Workshop on Computational / Experimental Approaches to Protein Defects in Human Disease, Rutgers University, April 2006.
88. University of North Carolina, Chapel Hill, Department of Chemistry, April 2006.
89. Duke University, Department of Biochemistry, April 2006.
90. UTMB, Galveston, TX, Sealy Center for Structural Biology, April 2006.
91. Johns Hopkins University, Department of Chemistry, March 2006.
92. Stanford University, Department of Chemistry, March 2006.
93. I2CAM Exploratory Workshop on Protein Aggregation and Amyloid Formation in Systemic and Neurodegenerative Diseases: Physical, Molecular, and Biological Approaches, EPFL, Lausanne, Switzerland, July 2005.
94. 18<sup>th</sup> Annual Gibbs Conference on Biothermodynamics, Carbondale IL, October 2004.
95. University of Iowa, Department of Chemistry, Iowa City, May 2004.
96. Washington University, Alzheimer's disease Research Center, April 2004.
97. Washington University, Department of Genetics, October 2002.
98. Washington University, Alzheimer's disease Research Center, December 2002.
99. Washington University, Biophysical Evening Seminar Series, December 2001.

### TEACHING

1. *Physics of Biopolymers and Bioinspired Polymers*, BME 432 / 532, Fall 2019 onward. Audience: Seniors majoring in Biomedical Engineering, Chemical Engineering, and Mechanical Engineering and PhD students from different disciplines including BME, Biophysics, and Materials Science.
2. *Bioengineering Thermodynamics in Practice*, Laboratory Course, BME 329, Fall 2014 & 2015. Audience: Biomedical Engineering juniors.
3. *Bioengineering Thermodynamics*, BME 320B, Fall 2010 onward. Audience: Biomedical Engineering juniors.
4. *Bioengineering Thermodynamics Recitation*, BME 320A, Fall 2015 onward. Problem solving session. Audience: Biomedical Engineering juniors.
5. *Applied Mathematics for Biomedical Sciences*, Fall 2013. Audience: Graduate students in Biomedical Engineering & quantitative programs in the Division of Biology & Biomedical Sciences.
6. *Chemical Thermodynamics*, ChemE 320 / BME 320, Fall 2008 and 2009. Audience: Chemical Engineering sophomores and Biomedical Engineering juniors.
7. *Biomedical Engineering Design*, BME 401, Fall 2007. Co-course master for the capstone senior design course. Audience: Biomedical Engineering seniors, and graduate students in BME and Molecular Biophysics.
8. *Principles of Protein Structure*, BME 461, Every Fall starting Fall 2003. Audience: Seniors and graduate students in Biomedical Engineering.
9. *Introduction to Biomolecular Statistical Thermodynamics*, BME 531, Graduate Level, Every Spring, starting Spring 2002. Audience: Graduate students in Biomedical Engineering, Chemical Engineering, and Molecular Biophysics. Last time course was taught: Spring 2007.

10. *Modeling Biomolecular Systems, Part II, BME 540*, Fall 2004. Audience: Graduate students in Biomedical Engineering, Chemical Engineering, Molecular Biophysics, and Computational Biology.
11. *Quantitative physiology, part II, BME 301B, Four Lectures on Applications of control theory in modeling physiological systems*, Spring 2003 – Spring 2005. Audience: Juniors majoring in Biomedical Engineering.
12. *Quantitative physiology, part II, BME 301B, Two Lectures on the Quantitative aspects of Antigen-Antibody Interactions and their role in control and regulation of immune response*, Spring 2006. Audience: Juniors majoring in Biomedical Engineering.

## **LEADERSHIP EXPERIENCE**

### ***Leadership Within Washington University***

#### **Center for Biological Systems Engineering**

I co-chaired the strategic planning committee commissioned by the Dean of the School of Engineering & Applied Sciences. The goal was to craft a strategic plan for the next 5-10 year period for the school.

I am the founding Director of the Center for Biological Systems Engineering (CBSE). This center is the home of network biology at Washington University and has incorporated researchers from the School of Engineering & Applied Science (SEAS) and the School of Medicine. These researchers deploy multiscale, systems-based approaches to model, predict and design functions of biological systems that result from the integration of signals and responses of biomolecular and cellular networks.

I led a cluster search to recruit suitable investigators to populate the CBSE. The search was successful in recruiting three new investigators, two by way primary appointments in Biomedical Engineering (BME), and one with primary appointment in Pathology & Immunology (P&I). Additionally, I recruited out to four kindred scientists from BME and P&I with expertise in multiscale approaches to challenging problems in biomedical science. Within a short period, I assembled a talented group of eight investigators, six of whom are assistant professors.

Research interests within the center span a range of topics including modeling protein self-assembly and homeostasis networks, reverse engineering networks controlled by post-translational modifications, utilizing and adapting novel sequencing methods to interrogate the organization of small RNA networks, chemical informatics geared toward prospecting for and repurposing novel small molecules, de novo design of protein interaction networks, and developing advanced imaging modalities for non-invasive interrogations of biological tissues.

#### **Center for High Performance Computing**

I am currently the co-director, with Dr. Fred Prior, of the Center for High Performance Computing (CHPC) at Washington University. My close involvement with the CHPC has contributed to a fundamental improvement of the computational environment at Washington University, which was essentially non-existent a few years ago. This effort has required several delicate interactions and management of important issues, and I have learned to become proficient in cultivating inter-personal relationships that enable rather than compromise the success of projects.

#### **Department of Biomedical Engineering**

I have contributed mainly as a model citizen to the curricular and research programs within the BME department. In addition to participating on key departmental committees, I took the lead on two specific curricular matters. I took over the teaching of Thermodynamics for our BME undergraduates and redesigned this course to emphasize the importance of thermodynamics concepts across the molecular, cellular, and physiological scales. This initiative helped improve the synergy between thermodynamics and the two quantitative physiology courses that are taught in the department. I added a focused problem-solving module to this course. I proposed the idea of developing a new laboratory module to go along with this course, and this has come to fruition following the recruitment of a suitably qualified instructor to handle these duties. My efforts in Bioengineering Thermodynamics help illustrate my ability to craft a vision and see the vision through to fruition through earnest effort and refinement of the vision by incorporating critical feedback.

I have brought a similar approach to bear on the topic of developing a new applied mathematics course for biomedical sciences. I am currently teaching this course that I developed from scratch. It is being designed to provide a direct conduit from concept to application of mathematics in biology and biomedical sciences. It is the only course of its kind at Washington University and this has arisen entirely through my initiative.

From a research standpoint, I have been actively involved in faculty recruitment within BME, have chaired and been a member of search committees, and I was the primary author of the white paper that was crafted roughly two years ago for the future research mission of the BME department. In crafting this white paper, I worked closely with a key senior colleague in the department, and actively solicited and incorporated the inputs from all of my BME colleagues, thus showcasing my ability to be both visionary and collaborative.

## SERVICE

### **Professional activities**

1. Reviewer of manuscripts for *Biochemical Journal*, *Biochemistry*, *Biomacromolecules*, *Biophysical Journal*, *Biophysical Chemistry*, *Cell*, *Cell Reports*, *eLife*, *Journal of Chemical Physics*, *Journal of Molecular Biology*, *Journal of Physical Chemistry B*, *Structure*, *Journal of the American Chemical Society*, *Journal of Chemical Theory and Computation*, *Journal of Computational Chemistry*, *Nature*, *Proceedings of the National Academy of Sciences USA*, *PLoS: Computational Biology*, *PLoS One*, *Science*, *Science Signaling*, *Structure*.
2. Panelist for review of National Science Foundation SBIR grants, Bioinformatics.
3. Panelist and reviewer of grants for National Science Foundation, Molecular & Cellular Biophysics.
4. Ad hoc reviewer for National Science Foundation, Biotechnology.
5. Grant Reviewer for Alzheimer's Disease Research Center, Washington University
6. Grant Reviewer for HighQ Foundation.
7. Co-author and contributor to Statement of Significance Petition to form a new Intrinsically Disordered Protein subgroup within the Biophysical Society.
8. Primary organizer of 1<sup>st</sup> International ICAM workshop on Multiscale Interactions and Dynamics in Complex Biological Systems, Washington University in Saint Louis, May 27-29, 2006.

9. Co-organizer for 20<sup>th</sup> Annual Gibbs Meeting on Biothermodynamics, October 7-10, 2006, Carbondale, IL.